

Crop Protection PLSS 6344

Lecture 11: **I**ntegrated **P**est **M**anagement “Sampling”

Overview

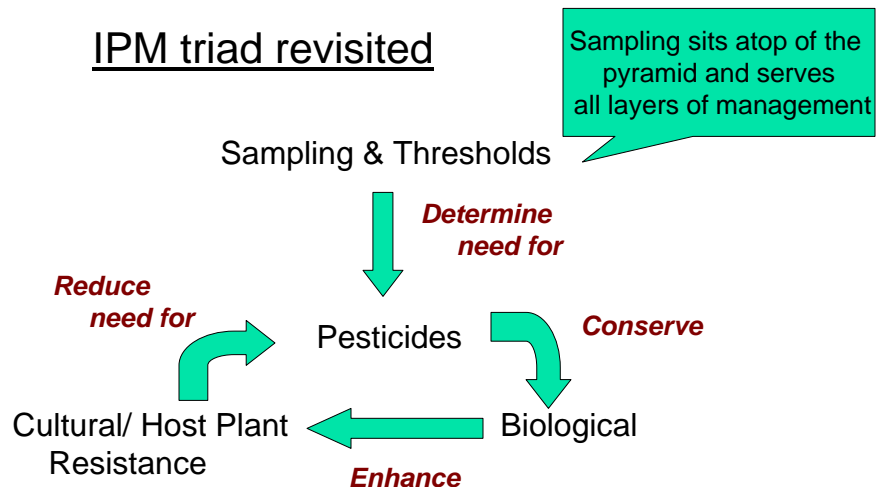
- I. Introduction (Estimates vs. Exact #; Sampling Program)
- II. Biological vs. Statistical Populations
- III. Types of sampling programs
- IV. Preliminary sampling Program
- V. Sampling Design
- VI. Sequential sampling

Lecture objectives

- Understand the concept of sampling and its importance in pest management
- Theoretical and practical aspects of sampling
- Students will learn how to initiate basic sampling plans for a given arthropod (insect) pest

Importance of Sampling in IPM

IPM triad revisited



I. Introduction

- **Sampling** = Collection of field observations to obtain a reliable estimate of the size of the arthropod population in a given area (e.g. # stalkborers in a corn field; boll weevil population in cotton field; # mosquitoes in a pond)
- **Census vs. Sampling** (Nearly impossible to count all inds. of an arthropod population in a given agroecosystem, sampling is thus **necessary**)
- 3 estimates of arthropod populations:
 - Absolute
 - Relative
 - Population Indices

I. Introduction

■ Absolute vs. Relative Estimates

- **Absolute** = all arthropods within the sample unit (difficult & intense. Often used in research but not in IPM systems)
 - **Extraction** (Vacuum sampler, sieving for non-mobile stages)
 - **Direct counting** (Vegetation removal for direct counting of thrips on onion, stemborers from cane splitting-easy w/large insects, but very difficult at higher density)
 - **Final absolute estimate** = $(\text{counts/ unit}) \times (\text{Total sample units/acre})$

■ Characteristics of Absolute numbers

Want a best measure of density as possible

- **Accuracy** - closeness of a measure to the true value
- **Precision** - closeness of repeated measures to each other
- **Bias** - degree of systematic error in estimating mean
- **Fidelity** - accuracy with which population estimates over time reflect actual changes in population numbers

I. Introduction

■ Absolute vs. Relative Estimates

- **Relative** = No direct relation to the area, depends on technique--Use for comparison of pops.
 - **Sweep net** (E.g. # of leafhoppers per sweep)
 - **Trap** (yellow or blue sticky card - Pheromone traps, can be sex biased)
 - **Plant damage (Pop. Indices)** (Damage scoring or rating, disease presence)

- **Characteristics of relative numbers**
 - Most often used in IPM
 - Cost efficient
 - Easy in most cases (but can be difficult in others)
 - Higher variance; may be difficult to interpret
 - Relative estimates can be converted into absolute estimates (regression analysis, experimentation etc..)

I. Introduction

■ Sampling Program

- **Sampling technique** = method used to collect single sample

- **Sampling Program** = How a sampling technique is used
 - Number of samples
 - Spatial and temporal patterns, etc...

- **Sampling Design** = Processes of establishing a sampling program

II. Biological vs. Statistical

- **Biological population** = actual # of arthropod in a given area (E.g. # of SCB in a sugarcane field: this # can be up to 40,000/ha. Impossible to count so biological pop. is often unknown) **[Rightly said it is a known unknown!]**
- **Statistical population** = estimate of the biological population determined by sampling a proportion of the biological population (In the eg above, if SCB is counted on 10 plants sampled per field, statistical population = # larvae on those 10 plants)
 - Estimates from statistical pop. obtained from sampling, needs to be summarized
 - Descriptive statistics (most common to describe arthropod pop. characteristics)

II. Biological vs. Statistical

■ Descriptive statistics

1. **Mean** = arithmetic average of the sample numbers

$$\bar{x} = \frac{x_1 + x_2 + x_3 + \dots + x_N}{N}$$

N = number of samples and \bar{x} = mean

- E.g. If SCB counts in 10 sampling units (plants) were: 4, 0, 12, 9, 18, 21, 7, 11, 12, 15 what would be the mean?
- Average of “several means” of this type from subsets will give m , the best estimate of population size.

The mean is the most important characteristic of an arthropod population-central tendency-but does not tell all! Other statistics are needed.

II. Biological vs. Statistical

- **Descriptive statistics-Variation (numerical differences among samples)**
 2. **Range** = Largest value - smallest value (E.g. 21 - 0)
 3. **Variance** = spread of individual measurements of the statistical population
 4. **Standard Deviation (SD or s)** = Average of deviation of sample counts from the mean value - SD is the square root of variance
 5. **Standard Error** = Expected spread of the sample means is repeated samples of the same size are taken from the population. **Magnitude decreases as # samples increased!**

II. Biological vs. Statistical

- **Descriptive statistics-Variation**
 - Statistical programs on PC or Handheld calculators can perform most of these calculations

Sample Number (Ni)	Sample Count (X)	X ²
1	4	16
2	0	0
3	12	144
4	9	81
5	18	324
6	21	441
7	7	49
8	11	121
9	12	144
10	15	225
SUM	109	1545
Mean	$x = 109/10 = 10.9$	

$$s = \sqrt{\frac{1}{N-1} (SUMX^2 - \frac{(SUMX)^2}{N})} = 6.30, V = s^2 = 37.7$$

$$SE = s / \sqrt{N} = 6.3 / \sqrt{10} = 2.0$$

III. Types of Sampling

Two main types of sampling Programs

A. Extensive

- Conducted over broad area
- To predict damaging levels of pops.
- Area-wide surveys
- Pattern of pop. dynamics (flight of Monarch butterfly)
- Coarse estimate-few samples are usually taken, one stage sample, systematic pattern

III. Types of Sampling

Two main types of Sampling Programs

B. Intensive

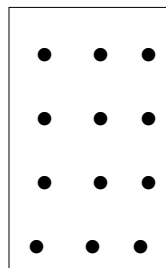
- Analytical-used for research (frequent observation of species in a small area-GWSS egg parasitism in south TX)
- Used for life table, mortality, natality, migrations
- Often several stages sampled
- Requires higher precision level

IV. Preliminary Sampling Program

- **A. Used to start a program, estimate workable program (What – Where – When - How?)**
 1. Try different techniques (Visual counts, mass trapping, damage estimate)
 2. Knowledge of basic attributes of arthropod biology required
 - **What is it?** Proper and accurate identification is the first step
 - **What is the life history** (where does the insect pest feed on a plant, how many generations a year, where are the eggs deposited?)
 3. Activity periods
 - **nocturnal vs. diurnal patterns; cool mornings vs host afternoons** (E.g., Lep. Moths; Alate aphids fly primarily during morning and late afternoon hours)

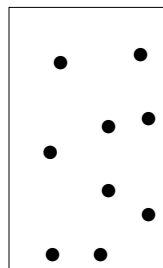
IV. Preliminary Sampling Program

- **Used to start a program, estimate workable program**
 4. Dispersal pattern (spatial arrangement)
 - **three basic patterns: aggregated, random and regular**



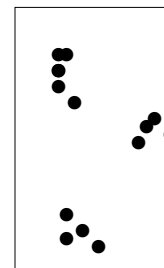
Regular

$$\sigma^2 < \mu$$



Random

$$\sigma^2 = \mu$$



Clumped

$$\sigma^2 > \mu$$

IV. Preliminary Sampling Program

- **Used to start a program, estimate workable program**

4. **Dispersal pattern:** Before making any statistical inference, test the relationship of arthropod distribution to known appropriate statistical distribution:

- > **Normal** = theoretical distribution wherein variation is random, mean and variance are independent
- > **Poisson series** = distribution of data with low mean (high proportion of 0)
- > **Bionomial** (Presence/Absence data)
- > **Negative bionomial** = variance is dependent upon the mean (clumped distribution)

Compute the mean and variance and compare them using established statistics

IV. Preliminary Sampling Program

- **Poisson: test of conformity**

Poisson probability: $P(n) = e^{-\bar{x}} \left(\frac{\bar{x}^n}{n!} \right)$

- **Aggregation index** = variance to mean ratio ($I_D = s^2 / \bar{x}$); this ratio is compared to 1 using chi-square test

$I_D < 1 \equiv$ regular; $I_D = 1 \equiv$ random and $I_D > 1 \equiv$ aggregated

- **Taylor's Power Law** = exponential function to describe the relationship between variance and mean

$$s^2 = a \bar{x}^b$$

Log-transformation gives: $\log(s^2) = \log(a) + b \log(\bar{x})$

b index of aggregation, $b < 1 \equiv$ regular, $b = 1 \equiv$ random, and

$b > 1 \equiv$ aggregated distribution

IV. Preliminary Sampling Program

Example: Data on EB on maize compared to Poisson distribution

Number of EB/cob	Observed frequency	Prob. Poisson	Expected freq.
0	1162	0.32	735.6
1	427	0.36	838.6
2	391	0.21	478
3	142	0.08	181.6
4	82	0.02	51.8
5	53	0.01	11.8
6	25	0.00	2.2
7	12	0.00	0.4
8	6	0.00	0.1

$$e^{-x} \left(\frac{x^n}{n!} \right)$$

N = 2300

Mean = 1.14; variance = 3.53

Chi-square = $\sum[(O_i - E_i)^2 / E_i] = 1,919.6$; P < 0.0001

Sétamou et al. 2000

IV. Preliminary Sampling Program

- **EB data analyzed with I_D**

I_D variance/mean = $3.53/1.14 = 3.10$

chi-square = $(n-1) \times I_D = 59.02 > 1$; n = 140

What is the Conclusion??

- **EB data analyzed TPL**

$\log(\text{variance}) = 1.26 + 1.47 \cdot \log(\text{mean})$

b = 1.47 > 1, t = 24.64, df = 130

What is the Conclusion??

IV. Preliminary Sampling Program

- **A. Used to start a program, estimate workable program**
 5. Determine sampling universe (broad area from which sample should be drawn)
 6. Trial and error (experiment to find most acceptable procedures)
- **B. Objectives**
 - Reduce costs
 - Plan sampling program with low variances

V. Sampling Design

- **A. Elements of the Sampling Program (6 key elements)**
 1. Sampling techniques
 2. Number of samples (per unit habitat)
 3. Sampling Unit (size, selection, habitat)
 4. Pattern of sampling
 5. Timing of sampling
 6. Sampling Interval

V. Sampling Design

- A. Elements of the Sampling Program

1. Sampling techniques
2. **Number of samples** (per unit habitat) : Compromise between accuracy and cost

Continuous: $N = [(t \times s)/(D \times \bar{x})]^2$

Bionomial: $N =$

N= number of samples required; t = Student's t-value;

s = standard deviation; \bar{x} = mean; D = required level of precision

D = 10% or 0.10 for research

D = 25% or 0.25 in IPM

V. Sampling Design

- A. Elements of the Sampling Program

1. Sampling techniques
2. **Number of samples**

Example: EB # required for 25% precision level

mean= 1.14; variance = 3.53, s = 1.88

$$N = [(1.96 \times 1.88)/(0.25 \times 1.14)]^2 = 169$$

For 10%

$$N = [(1.96 \times 1.88)/(0.10 \times 1.14)] = 1045$$

V. Sampling Design

- **A. Elements of the Sampling Program**
 1. Sampling techniques
 2. Number of Samples
 3. **Sampling Unit** (land area, whole plant, plant parts, etc..
 - a. Equal chance of selection
 - b. Stability (same plant growth stage)
 - c. Proportion of arthropods within each unit kept constant
 - d. Can convert into unit area
 - e. Balance between cost and accuracy
 - f. Sampling unit easy to be determined in the field

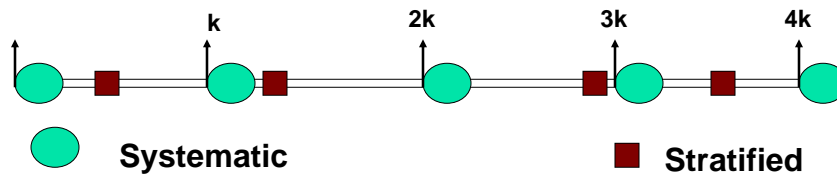
V. Sampling Design

- **A. Elements of the Sampling Program**
 1. Sampling techniques
 2. Number of Samples
 3. Sampling Unit
 4. **Sampling Pattern**
 - a. Depend on objectives of program
 - b. Types of sampling patterns
 - i. Unrestricted random (equal chance, not variance efficient)
 - ii. Stratified random (subdivision into strata, more efficient and very common in IPM)
 - iii. Systematic (equal distance sampling, analysis difficult)

V. Sampling Design

■ A. Elements of the Sampling Program

1. Sampling techniques
2. Number of Samples
3. Sampling Unit
4. Sampling Pattern



V. Sampling Design

■ A. Elements of the Sampling Program

1. Sampling techniques
2. Number of Samples
3. Sampling Unit
4. Sampling Pattern
5. Timing
 - Consider Temperature effects (DD to predict arthropod occurrence)
 - For unknown spp., sample regularly until optimum time is detected
6. Sampling Interval
 - Depend on generation time (short for fast developing pest sp.: 2-3 d for aphids; stemborers 1 to 2 weeks)
 - Consider the environment!

VI. Sequential Sampling

- **Reduce costs while maintaining precision**
- **Sampler make decision:**
 - Numbers too low stop sampling
 - Numbers very high and take treatment action
 - Numbers in between threshold, too close to call, recheck later
- **Method:**
 - Threshold lines predefined (lower and higher)
 - Compare successive sample tally to those limits

VI. Sequential Sampling

Sample no.	Don't treat	Tally 1	Tally 2	Tally 3	Treat
1	-	0	1	1	-
2	-	1	1	2	-
3	-	2	2	2	-
4	-	3	2	3	-
5	-	3	3	4	-
6	3	4	4	5	7
7	4	5	5	5	7
8	4	5	5	5	8
9	4	6	5	6	9
10	5	6	6	7	9
11	5	7	6	7	10
12	6	8	6	8	10
13	6	9		8	11
14	7	10		8	11
15	7	10		9	12
16	8	11		10	12
17	8	12		10	13
18	9	13		11	13
19	9			11	14
20	9			11	15

Treat Don't Recheck

Summary: Sampling Programs

- Sampling has been defined
- Biological pop. unknown, Statistical pop. used to make inference on Biological
- Factors to be considered in sampling, Sample size
- Different sampling patterns

“ The sampling program is a mixture of art, science, and drudgery; art and science are in the sampling design”

R. F. Morris